



## **Professional Development**





## **PV92 PCR Informatics Kit:** Where did you get those GENES?







#### Chromosome 16: PV92 PCR Informatics Kit

### Instructors



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#### Why Teach Polymerase Chain Reaction (PCR)?



- Powerful teaching tool
- Real-world connections
- Link to careers and industry
- Tangible results
- Laboratory extensions
- Standards-based





#### Scientific Inquiry Chemistry of Life Use of PCR and DNA gel DNA extraction techniques electrophoresis in DNA profiling DNA replication and PCR. Use of positive experimental DNA structure, function, and controls. chemistry Use of bioinformatics databases Chemical properties of biological Apply the Hardy-Weinberg molecules equation to student data Genetics Evolution Mendelian genetics Function of genetic diversity Homozygous vs. heterozygous Genetic variation in the human. alleles genòme Inheritance of dimorphic loci Selective advantages of Genetics of noncoding DNA heterozygous alleles Short repetitive interspersed elements (SINEs) Environmental and Health Science Cell and Molecular Biology Eukaryotic cell structure and Bioinformatics to compare class organization data to worldwide population Tissue types for biological data sampling Molecular genetics to study human migration patterns Role, place, limits, and possibilities of science and technology





#### Chromosome 16: PV92 PCR Informatics

## **Kit Advantages**



- Aligns with AP Biology AP Lab 8
- Extract genomic DNA and amplify student samples
- Introduce the polymerase chain reaction (PCR)
- Apply PCR to population genetics
- Directly measure human diversity at the molecular level
- Compare results to online data
- Sufficient materials for 8 student workstations
- Complete activity in three 45 minute sessions





#### Chromosome 16: PV92 PCR Informatics Kit Workshop Timeline

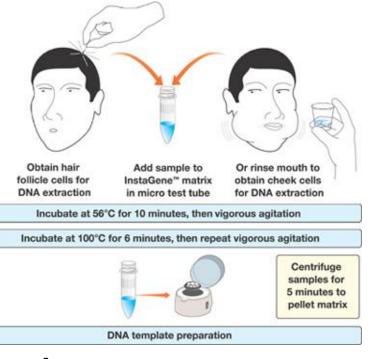


- Introduction
- Extract genomic DNA and prepare samples for PCR
- Cycle samples
- Agarose gel analysis
- Hardy-Weinberg analysis
- Bioinformatics

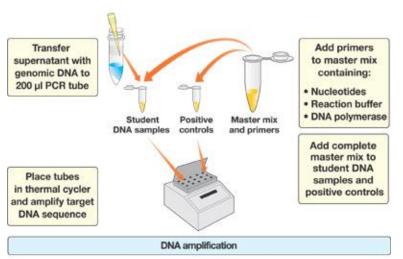




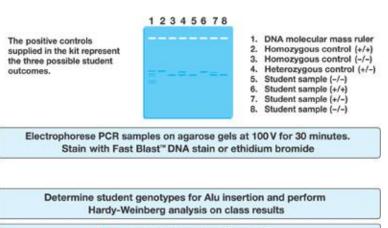
#### **Chromosome 16 PV92 PCR Procedure Overview**



Day 1



#### Day 2



Extension: Web-based bioinformatics

#### Day 3







- DNA replication gone crazy in a test tube!
- Makes millions of copies of a target sequence from template DNA
- Uses heat-resistant *Taq* polymerase from *Thermus aquaticus*





### Laboratory Quick Guide

speed for 2 minutes. When the centrifuge has completely stopped, remove your tube. You should see a match-head sized pellet of whitish cells at the bottom of the tube. If you don't see a pellet of this size, decant the saine, rem liv your tube with more of your oral rinse, and repeat the spin.
<ul> <li>test tube (NOT the screwcap tube) with your initials. If a P-1000 micropipel is nd available, carefully pour ~1 mi of your saline rinse into your micro test tube (use the graduations on the side of the micro test tube to estimate 1 mi).</li> <li>Spin your tube in a balanced centrifuge at full speed for 2 minutes. When the centrifuge has completely stopped, remove your tube. You should see a match-head sized pellet of whitish cells at the bottom of the tube. If you con't see a pellet of this size, decant the salin, refully our tube with more of your oral rinse, and repeat the spin.</li> <li>After pelleting your cells, pour off the saline. Being careful not to lose your pellet, biot your</li> </ul>
completely stopped, remove your tube. You should see a match-head sized pellet of whitish cells at the bottom of the tube. If you don't see a pellet of this size, decant the saine, refli your tube with more of your oral rinse, and repeat the spin. 5. After pelleting your cells, pour off the saine. Being careful not to lose your pellet, biot your
Being careful not to lose your pellet, blot your
for a small amount of sallne (< 50 µl, about the same size as your pellet) to remain in the bottom of the tube.
<ol> <li>Resuspend the pellet by vortexing or floking the tube so that no clumps of cells remain.</li> </ol>
7. Using a 2-20 µl adjustable-volume micropipet set to 20 µl, transfer all of your resuspended oelis to the screwcap tube containing InstaGene.
<ol> <li>Screw the cap tightly on the tube. Shake or vortex to mix the tube contents.</li> </ol>





### Micropipet Use



- **1.** Twist dial to desired volume
- 2. Pick up pipet tip
- 3. Press plunger to first, soft stop
- 4. Insert pipet tip into solution to be transferred
- 5. Slowly release plunger to retrieve liquid
- 6. Move pipet tip into desired tube
- 7. Press plunger past first stop to second, hard stop to transfer liquid



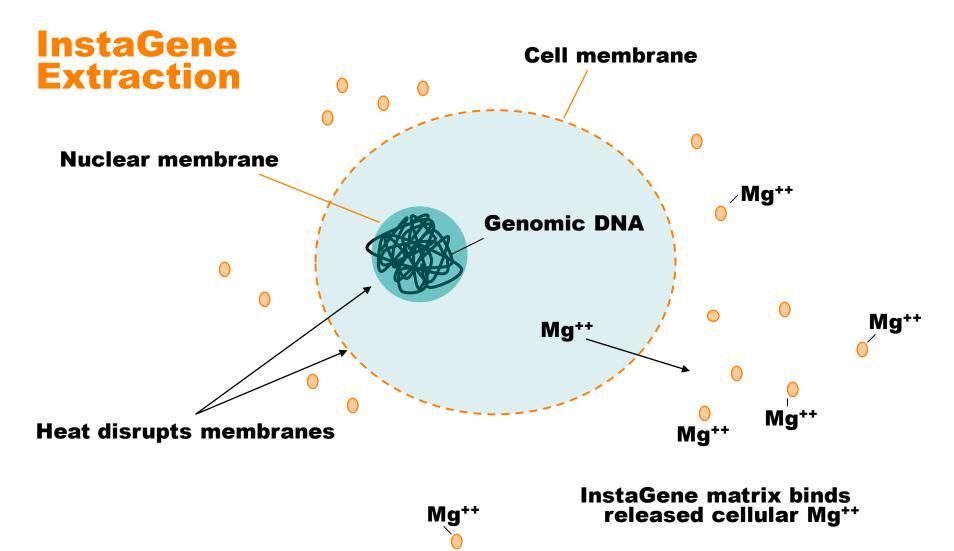


### **Protocol Highlights** Genomic DNA Extraction

- InstaGene<sup>™</sup> Chelex<sup>®</sup> cation exchange resin; binds cellular magnesium ions
- 56°C loosens connective tissue and inactivates DNAses
- 100°C ruptures cell membranes and denatures proteins





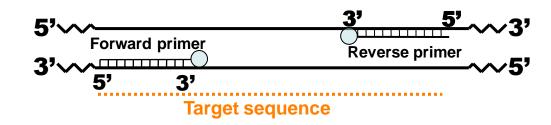






## What Is Needed for PCR?

- Template (the DNA you want to amplify for the study)
- Sequence-specific primers flanking the target sequence:



- Nucleotides (dATP, dCTP, dGTP, dTTP)
- Magnesium ions (enzyme cofactor)
- Buffer, containing salt
- *Taq* polymerase





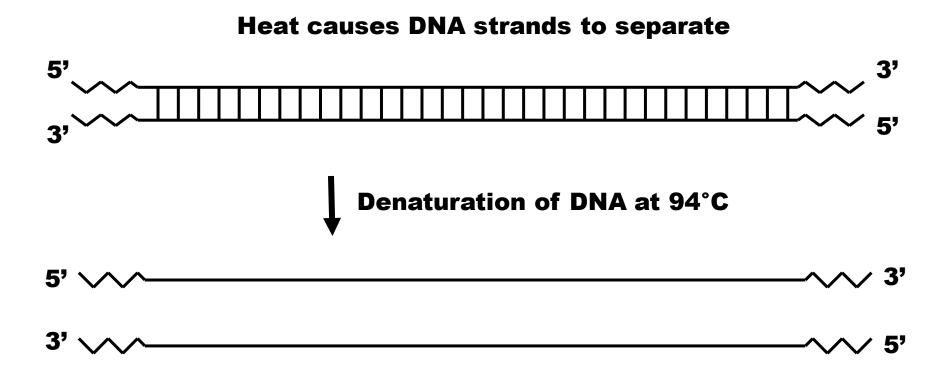
### How Does PCR Work?

- Heat (94°C) to denature DNA strands
- Cool (60°C) to anneal primers to template
- Warm (72°C) to activate *Taq* polymerase, which extends primers and replicates DNA
- Repeat multiple cycles





### Denaturing Template DNA

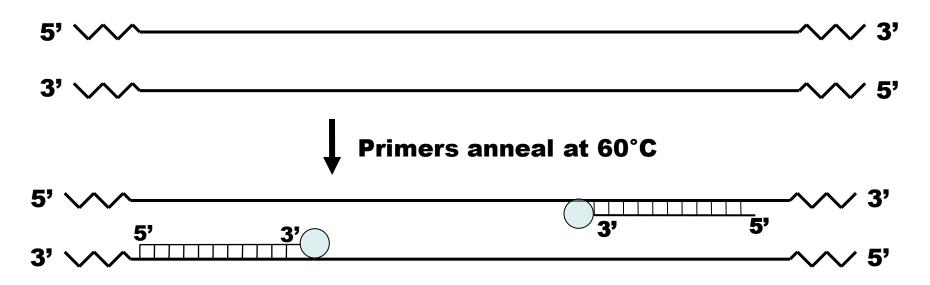






## Annealing Primers

- Primers bind to the template sequence
- *Taq* polymerase binds to double-stranded substrate

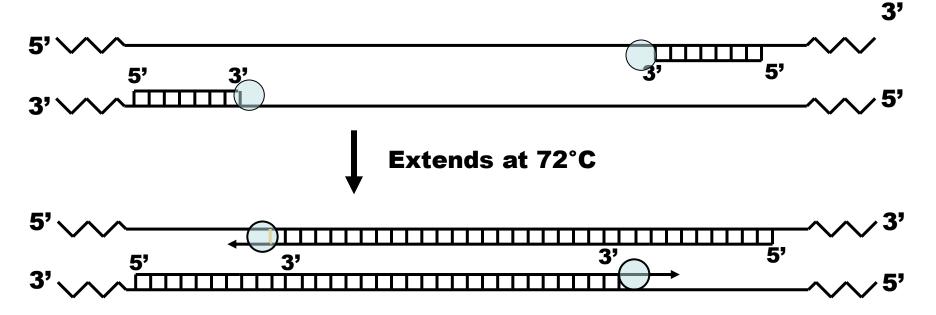






### *Taq* Polymerase Extends...

- *Taq* polymerase extends primer
- DNA is replicated

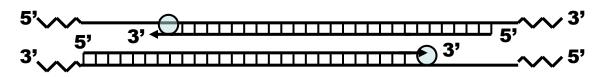




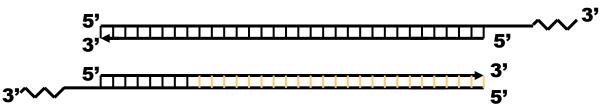


### Exact-length Target Product is Made in the Third Cycle











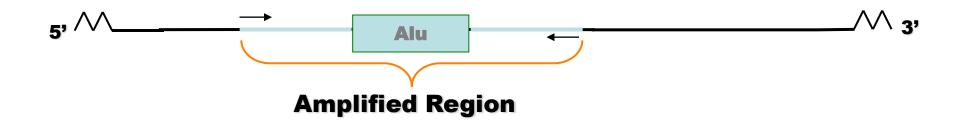






## The Target Sequence

- PV92 Alu insertion
- Located on Chromosome 16

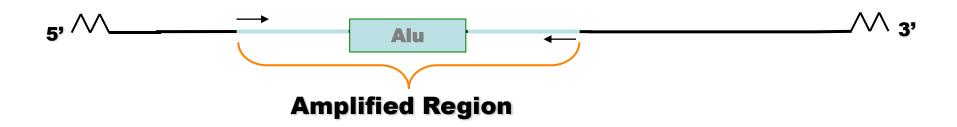






## PV92 *Alu* Insertion

- A member of *Alu* repeat family
- Human-specific Alu insertion
- Found in a non-coding region of your DNA
- Not diagnostic for any disease or disorder





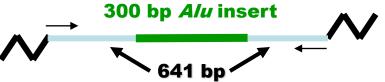


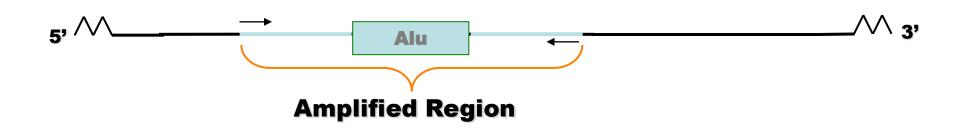
## **PCR Results**

 The PV92 Alu is dimorphic so there are two possible PCR products: 641 bp and 941 bp





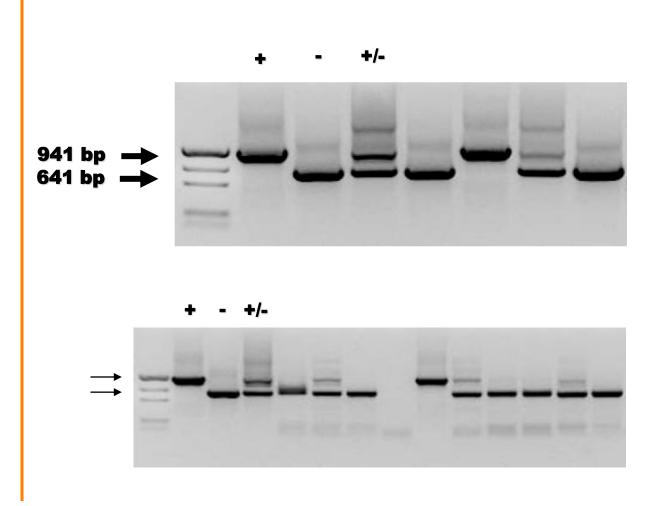








## Actual Alu PCR Results







## **Alu Repeats**

- Classified as SINEs (Short Interspersed Repetitive Element)
- Mobilized by an RNA polymerase-derived intermediate (retroposition)
- Approx. 500,000 *Alu* copies per haploid genome, representing about 5% of the genome
- Named for the *Alu* I restriction site within the element





### **Evolutionary Significance** of PV92 *Alu* Inserts

- Highly conserved
- Inserted in the last 1,000,000 years
- Genotypes (+/+, +/-, -/-)
- Used in population genetics, paternity analysis, and forensics





**Determination and analysis** of *Alu* Frequency in a population

- Amplify *Alu* insert from representative sample population
- Calculate the expected allelic and genotypic frequencies
- Perform Chi-square test





**Calculating** Observed Genotypic Frequencies

Genotype	+/+	+/	-/-	Total (N)
# of People	25	5	8	38
Observed Frequency	0.66	0.13	0.21	1.00

+/+ Genotypic frequency	=	Number with genotype Population total (N)
	=	<u>25</u> 38
	=	.66





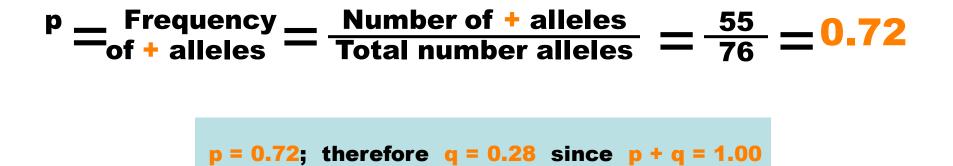
### **Calculating** Allelic Frequencies

Number of + alleles

25 individuals with two + alleles = 50 + alleles 5 individuals with one + allele = 5 + alleles Total = 55 + alleles

**Total number of alleles** 

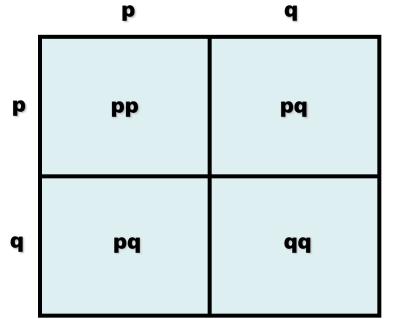
2N = 2(38) = 76







### *Alu* and Population Genetics



+/+ = p<sup>2</sup> +/- = 2pq -/- = q<sup>2</sup>

## Hardy-Weinberg Equation $p^2 + 2pq + q^2 = 1$



### Using the Hardy-Weinberg Equation to Determine Expected Genotypic Frequencies (p<sup>2</sup>, 2pq, q<sup>2</sup> values)

p <sup>2</sup>	+	<b>2pq</b>	+ q <sup>2</sup>	<b>_1.00</b>
(0.72)	) <sup>2</sup> +2	(0.72)(0.2	28) <b>+(0.2</b> 8	) <sup>2</sup> <b>_1.00</b>
0.52	+	0.40	+ 0.0	8 <u>=</u> 1.00
			- 0. 40	2 – 0 00









### Calculate Expected Numbers for Each Genotype

		Genotype frequency	Χ	Population total (N)	=	Expected number
0	<b>+/+</b> (p²)	0.52	Χ	38	=	20
Genotype	<b>+/</b> – (2pq)	0.40	X	38	=	15
)e	<b>—/—</b> (q <sup>2</sup> )	0.08	Χ	38	=	3





### Chi-Square Test

X<sup>2</sup> critical value (from statistics table) = 5.9

$$X^2 = \sum_{i=1}^{2} \frac{(Observed - Expected)^2}{Expected}$$

**16.25** is above 5.9 so the observed genotypic frequencies are not in genetic equilibrium

		Observed	Expected	<u>(O–E)²</u> E
G	+/+	<b>25</b>	20	1.25
Genotype	<b>+</b> /_	5	15	6.67
pe	_/_	8	3	8.33
X <sup>2</sup> = 16.25				16.25





## **Allele Server**

(1 of 17)

#### **Cold Springs Harbor Laboratory DNA Learning Center**

Web site: http://www.dnalc.org/





# **Allele Server**

(2 of 17)

#### Scroll through DNALC internet sites until BioServers Link appears



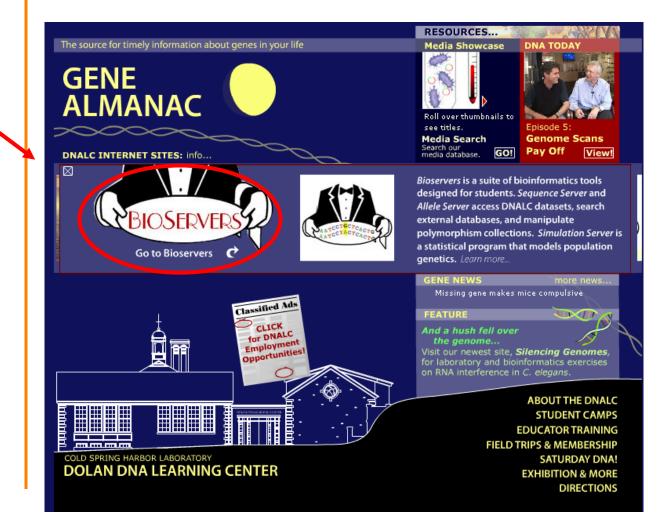




## **Allele Server**

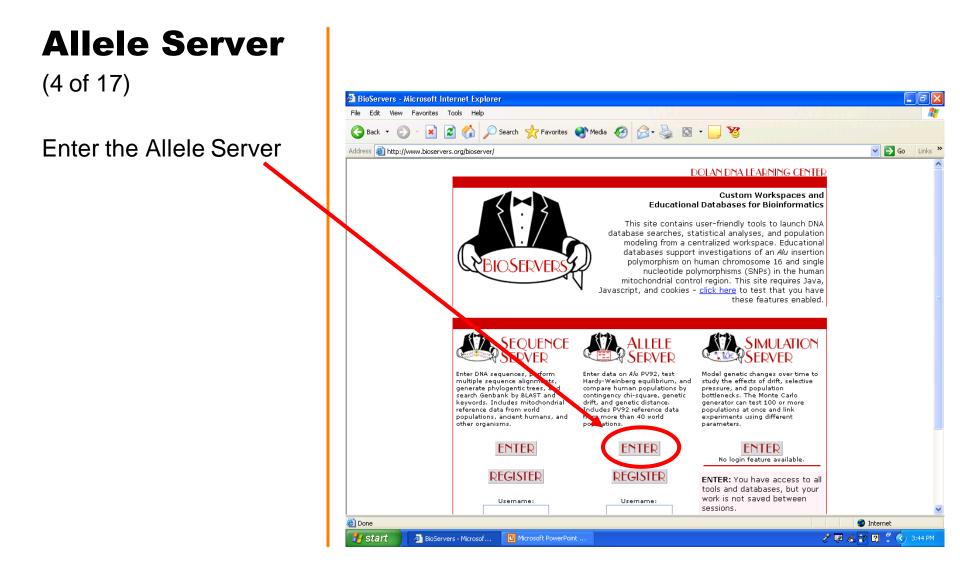
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**Click on Bioservers** 







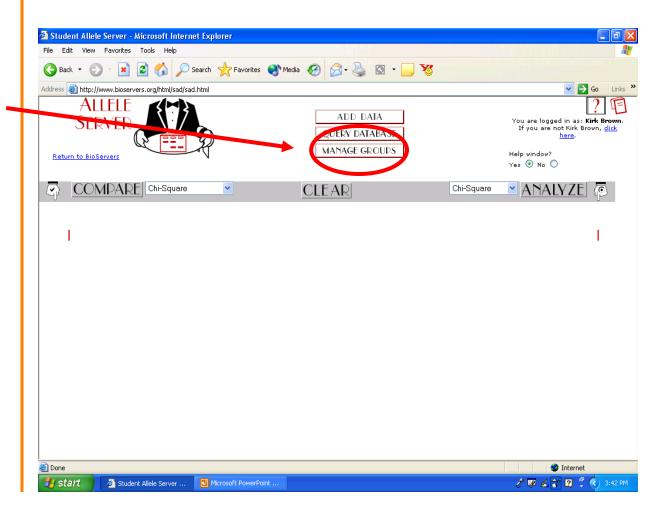






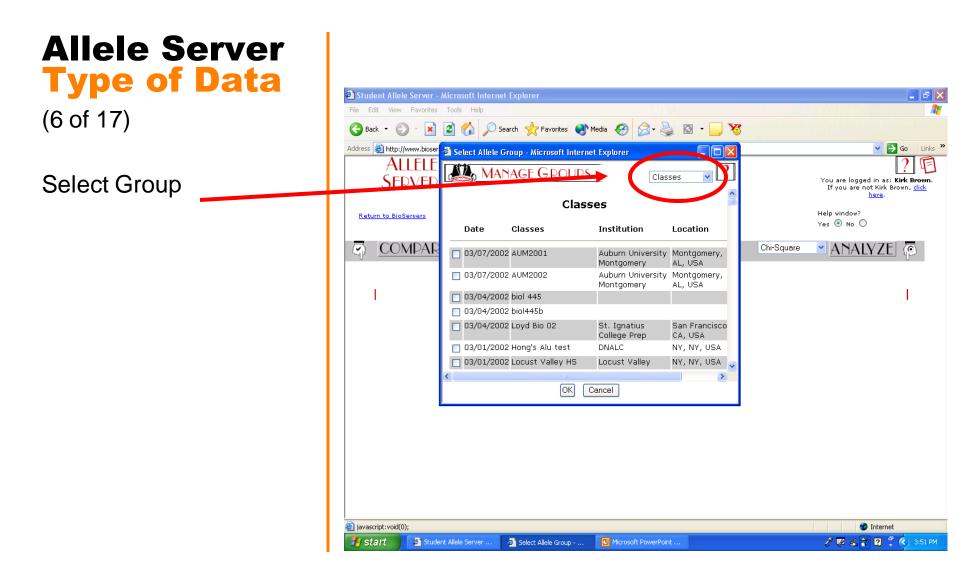
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Click on Manage Groups





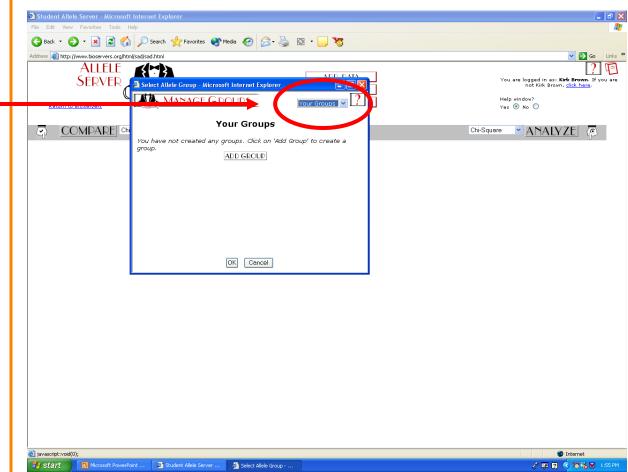








# Allele Server (7 of 17) Scroll Down to Select "Your Group"







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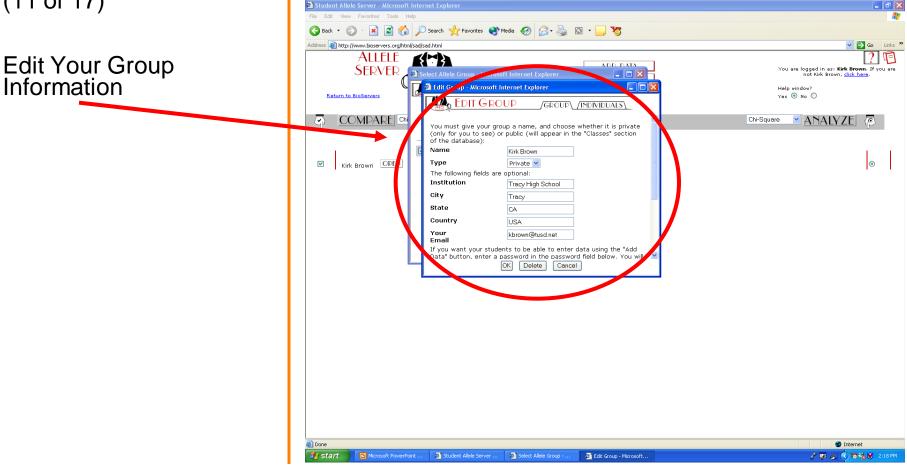
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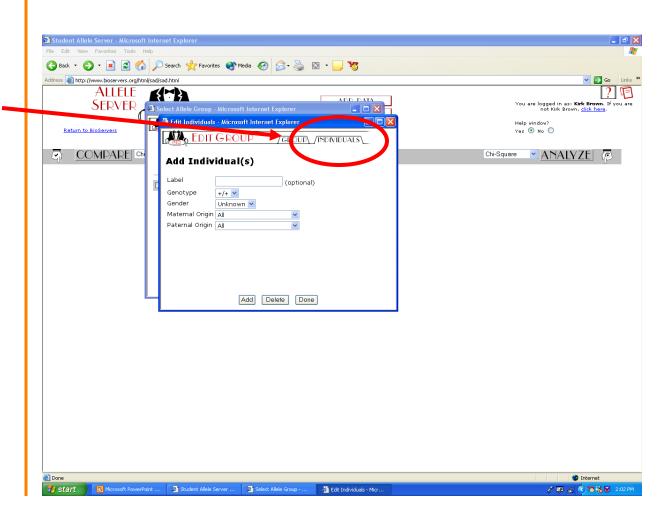






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Click on Individuals Tab





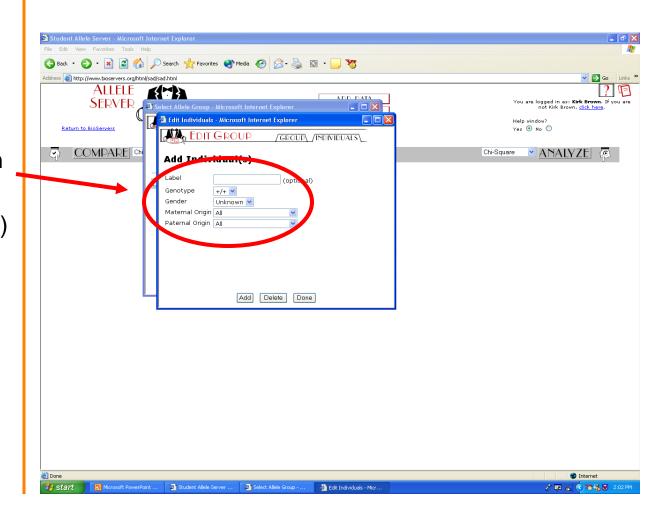


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Add Each Student's Information

Add as much information as possible:

- Genotype (+/+, +/-. -/-) Gender
- Personal Information

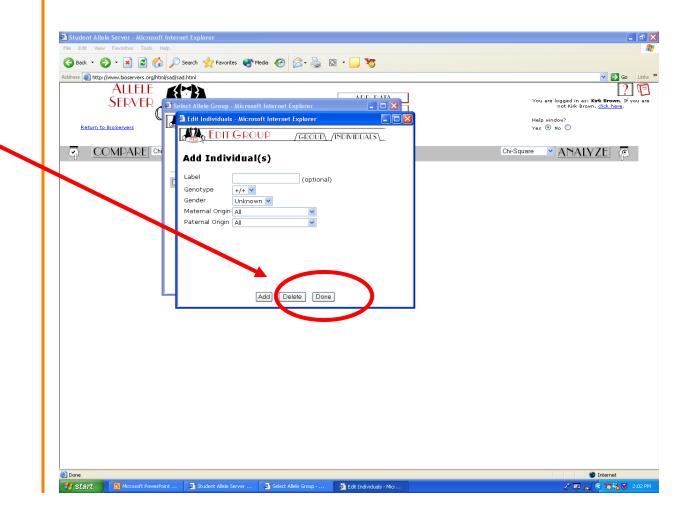






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**Click on Done** 

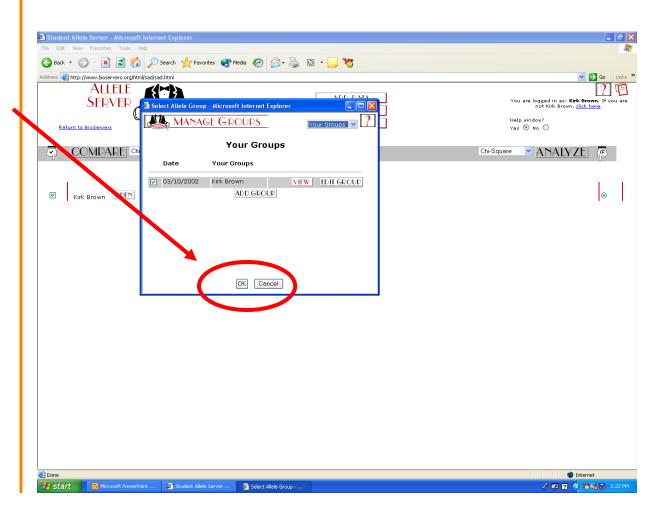






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Select and then Click OK

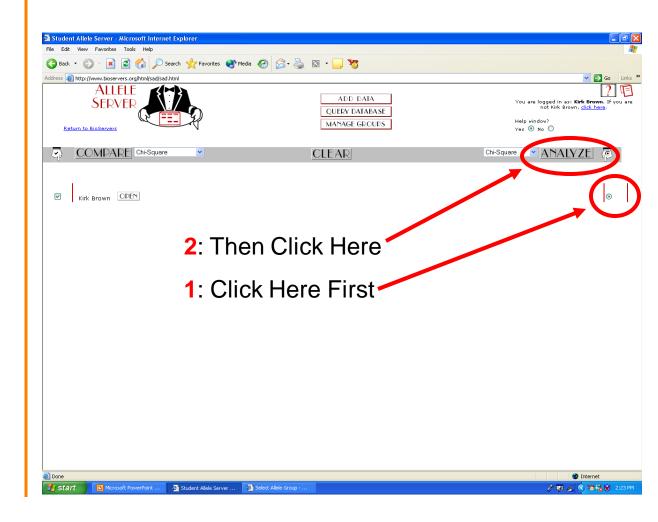






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Analyze Data

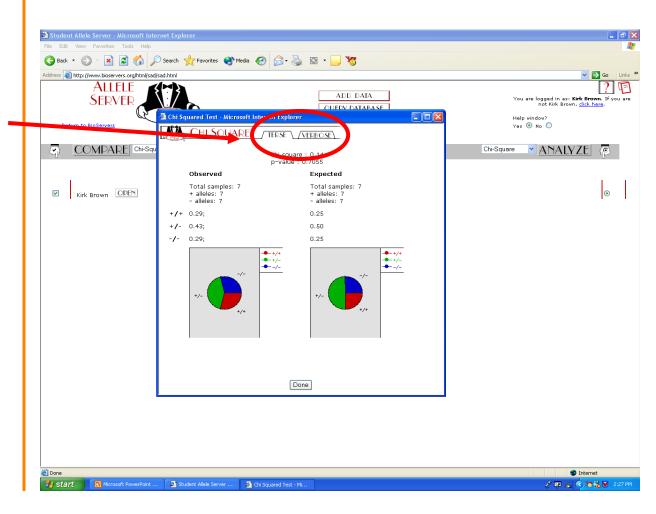






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Click on the Terse and Verbose Tabs to Review Data Results







#### **Extensions**

- Add each class separately and compare to see if the classes different from each other
- Compare your group to other existing groups
- Have students do manual calculations first and then compare to the computer generated version